Protocol for egg incubation

Planning for incubating the spotted wolffish eggs

All equipment for incubation should be procured before the spawning season approaches.

Here is how to plan for egg incubation:

- Prepare the incubation room, preferably a cool room to maintain a stable low temperature (for example, the cooling room at our faculty is set to 7.0 °C)
- Family hatchers (Figure 1) can be used to incubate the eggs
- Make sure that you have procured the required equipment to care for the eggs during the incubation period:
 - Red light headlamps (and batteries)
 - Disposable gloves
 - Coffee spoons (for water surface cleaning in the incubators)
 - Egg pipettes (6 or 7 mm)
 - Disinfectant (Buffodine) and related equipment (see disinfection protocol)



Figure 1: Family hatching chamber that can be used to incubate fertilized eggs from a single fish.

General guidelines for egg incubation

- Disinfect all equipment before, in between, and after every operation throughout the incubation period
- Salinity of water should be 33-34 % (we employed sea water pumped from 250 meter)
- Should use cold water filtered through 6 µm Faivre drum filters
- Optimum temperature is 5.0 °C, max 6.0 °C (our target was 5.5 °C)

- Preferred waterflow through the incubators is 160 liters/hour (flow rate can be increased, but the eggs should remain stationary)
- Oxygen saturation should be close to 100%
- The eggs are sensitive to light! Light exposure leads to increased mortality! Incubate in a dark, cold room
- Use red light while you tend the eggs or when you inspect their viability
- If possible, the eggs should be left undisturbed during the first 14-30 days after fertilization
- It should be noted that if you pick dead eggs during this period, you risk disturbing healthy eggs
- After the first period and if the eggs are of good quality, dead eggs should be removed only once in 2 weeks
- Check the eggs at least once a week and carefully remove the surface film from the water
- The eggs should be disinfected with buffodine once a week, until approximately 700 degree days
- Eggs should not be disinfected after 700 degree days to prevent premature hatching
- At approximately 280 degree days, the eggs would have developed visible eyes and would be more capable of tolerating handling stress
- If possible do not disturb the eggs as it can affect the survival rate

Transferring the eggs from the fertilization buckets into the family hatchers/incubators

- Be careful while you transfer the eggs from the fertilization bucket to the incubator
- Transfer the semen-egg mixture carefully and slowly, and the transfer should be done just above the water surface to avoid any drop
- Divide the eggs into batches, with a maximum of 1.5 L of eggs in each incubator
- After all eggs are transferred to the incubator, use either a utensil (no sharp edges), or your hand (use gloves) to distribute the eggs carefully as a thin layer in the incubator
- The eggs must not be disinfected until at least 12 hours post-fertilization

Identifying and discarding dead eggs

- Avoid disturbing healthy eggs while you remove the dead eggs
- Use an egg pipette to carefully remove loose dead/contaminated eggs from the incubated egg batch (Figure 2)

- If the dead/contaminated egg is not loose and cannot be removed easily, leave it for later picking in a few days
- Dead eggs should be discarded in an ensilage system



Figure 2: Removing dead eggs with a wide-diameter pipette.

Disinfecting the eggs

See the disinfecting protocol

Hatching - See the hatching protocol

Project is co-funded by the Kolarctic CBC program and Nordland County



Nord University Faculty of Biosciences and Aquaculture 8049 Bodø NORWAY

